THE EFFECT OF EIGHT WEEKS HIGH INTENSITY INTERVAL TRAINING (HIIT) ON SERUM AMOUNTS OF FGF21 AND IRISIN IN SEDENTARY OBESE WOMEN

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Abstract

Background & Aims: Transforming white adipose tissue to brown adipose tissue is considered a solution to overcome undesirable effects of obesity in human beings. Thus, the aim of this research was to investigate the effect of eight weeks of high intensity interval training (HIIT) on serum amounts of fibroblast growth factor 21 (FGF21) and Irisin in sedentary obese women.

Materials & Methods: Twenty sedentary obese women (with the average age of 30.15±2.96 and BMI of 30.34±1.27) were selected and randomly categorized into HIIT group and control group each consisting of 10 members. HIIT program was conducted for 8 weeks, 3 sessions in each week and with 90 percent of target heart rate. Blood samples were taken from both HIIT and control groups (while fasted) before and after the training to measure Irisin, FGF21 and lipid profile. The research data were analyzed by independent and dependent t-statistic tests in the significant level of P<0.05.

Results: The results of the statistical analysis showed HIIT program increased serum amounts of FGF21 and Irisin in sedentary obese women (P<0.05). Also, the results of independent t test showed there was a significant difference between HIIT group and control group in terms of FGF21 and Irisin amounts.

Conclusion: According to the results of this research, it seems HIIT program with increasing amounts of Irisin and FGF21 may lead to the transformation of white adipose tissue to brown adipose tissue and therefore, it has a key role in preventing obesity and its negative consequences.

Keywords: High Intensity Interval Training (HIIT), Irisin, FGF21, Sedentary obese women

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Introduction

The prevalence of obesity has highlighted the need for preventing its metabolic consequences through new treatment methods. It has recently been reported that transforming white adipose tissue (WAT, calorie storage) to brown adipose tissue (BAT, heat and calorie usage) is a solution for overcoming undesirable consequences of obesity in human beings (1, 2). White adipose tissue and brown adipose tissue play a key role in energy balance in the mammals. White adipose tissue is widely distributed in the body and is the first place in which fat is stored and burned. Brown adipose tissue is

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relatively few and is responsible for preserving body temperature through thermogenic adaptability (3,4). Heat production in brown adipose tissue is mostly related to adrenergic activity due to lipolysis and free fatty acids release through uncoupling protein 1 (UCP1) (5,6). Studies show white adipose tissue cells are capable to distinguish themselves from brown ones and vice versa. Once exposed to coldness, some of white adipose tissue cells change into brown ones to increase heat production. Being exposed to diet, brown adipose tissue changes to white adipose tissue to store energy (7).

Fibroblast growth factor 21 (FGF21) is amino acid hormone 181 from FGF family which functions as hyperglycemia, anti-fat, and heat producer (8). FGF21 is express in adipocytes, liver, skeletal muscles, and pancreas (9). Transcription of FGF21 is done in the liver under direct control of peroxisome proliferator-activated receptor alpha (PPARα) and by peroxisome proliferator-activated receptor gamma (PPAR γ) in adipose tissue (9). FGF21 has a positive and significant relation with UCP1 in BAT and through UCP1 and ACC1 leads to the activation and motivation of WAT (10). This process is adjusted by FGF21 through increasing the level of PGC-1α protein (11). Also, FGF21 adjusts energy hemostasis in adipocytes by activating AMP-activated protein kinase (AMPK) and sirtuin (SIRT). The result is an increase in the function of mitochondrial oxidative (11).

It is shown that FGF21 is a metabolic hormone which affects on energy balance and fat and glucose metabolism. The omission of FGF21 causes disturbance in glucose hemostasis and leads to fatness (12). The studies show that the overweight have more amount of FGF21 (13). However, it seems that FGF21 has anti-fat effects which lead to losing weight through the increase in using fat and reduction in fat mass (13). FGF21 is involved in metabolic disturbances pathogens such as insulin resistance, diabetes type 2, and fatty liver disease (14). It is shown that it can lead to natural revival of blood sugar level and triglyceride and improve insulin sensitivity regardless of weight in overweight and diabetic samples (13).

It is also believed that exercise may lead to improve metabolism of body by stimulating FGF21. Huwan Kim et al. (2013) showed that acute exercise for 30 minutes can increase FGF21 in both humans and animals. This increase is in tandem with an increase in free fatty acids and Glycerol (15). Also, Tanimura et al. (2016) showed that doing exercises with 75% VO2max for 60 minutes increased serum amounts of FGF21 (13). Other studies, however, don’t confirm these results. In this regard, Slusher et al. (2015) showed that 30 minutes of submaximal aerobic exercises creates less FGF21 in the obese people compared with people with natural weight (14).

Also, Kong et al. (2016) showed that 5 weeks of HIIT didn’t make any significant changes in FGF21 (14). Birjandi et al. (2016) showed that 6 weeks of HIIT didn’t make any significant changes in FGF21 in overweight and obese men (14). Since findings are not consistent, it is necessary to do more researches with longer durations and different intensities.

Irisin is a myokine derived from skeletal muscle which originates from fibronectin type III domain-containing protein 5 (FNDC5) and is dependent on PGC-1α for thermogenic activation of white adipose tissue and glycemic improvement (14). In fact, Irisin, through expression of UCP1, affects white adipose tissue and transforms it to brown adipose tissue and thus

4. acetyl-CoA carboxylase 1 (ACCI)

5. Peroxisome proliferator-activated receptor gamma coactivator 1-alpha (PGC-1α)
leads to the increase in calorie consumption and heat production. This, finally, results in losing weight and improving obesity (19, 20). Moreover, Irisin leads to the oxidation of fatty acids and glucose by activating AMPK which shows the role of Irisin in glucose hemostasis and fat (21). In this regard, Miyamoto et al (2015) showed that endurance training creates more Irisin in old men and reduces their stomach fat (22). Yang (2016) studied the effect of short-term and long-term swimming with moderate intensity in obese rats and showed that Irisin increased in both cases. The short-term exercise (8 weeks) resulted in gradual progression of obesity and long-term exercise (16 weeks) prevented its progression (23). Though a lot of studies support the stimulating effect of exercise on irisin level, Benedini (2017) showed that serum amounts of Irisin in professional and occasional athletes are the same as physically inactive individual (24). Given these results, it seems exercise type, exercise duration, and exercise intensity are influencing factors on Irisin secretion.

HIIT has recently been considered a proper exercise choice not only for athletes but for overweight and obese individuals who have limited time (25). HIIT uses exercise intensity as an effective factor instead of exercise volume. In exercises with low volume and high intensity, fat mass is lost in a more desirable way than consistent low-intensity exercises. For instance, it is shown that HIIT increases VO$_2$ max and reduces fat percentage and BMI (26). In general, different studies have been conducted in recent years on brown adipose tissue as an effective factor in energy hemostasis and preventing obesity. It has been shown that FGF21 and Irisin increase heat and energy consumption by transforming white adipose tissue to brown adipose tissue and activating UCP1 (27).

Nevertheless since a large number of studies deal with resistance and intensive exercises and HIIT exercises are less used, and given the time consuming nature of traditional resistance exercises, HIIT exercises can increase motivation of individuals to take part in the exercises. Also, given limited and contradictory results, more studies are required. As a result, this research aims to investigate the effect of eight weeks of high intensity interval training (HIIT) on serum amounts of FGF21 and Irisin in sedentary obese women.

**Materials and Methods**

The present research is quasi-experimental, pre-test and post-test with control group. The population of the research includes sedentary obese women in Naghade from 25 to 35 years old. Following the initial announcement for participation in the research, 40 sedentary obese women were selected. Then, 20 of them were randomly qualified to fill in the letter of satisfaction and healthy questionnaire and they were categorized in two groups: HIIT group (10 persons) and control group (10 persons).

The criteria for entering the research were as the following: BMI $\geq$ 30 kg/m$^2$, not having disease records, not being under medical treatment during the research, not having a regular exercise program. The criteria for leaving the research were: lack of interest, taking medicines and nutritional supplements, not taking part in exercise sessions regularly, and being hurt (14).

Participants were asked to take part in a briefing session. They filled in the letter of satisfaction and healthy questionnaire in this sessions and received necessary information for the following sessions. In the next session, they learned how to do HIIT exercise. Then, 5 cc blood was taken from their arm vein by a lab expert. Sampling was done 24 hours before exercise intervention while participants were fasted.

Next, the anthropometric measurements including height (by Wall stadiometer with 0.5 cm accuracy) and weight (by CAMRY, EF551BW digital scale with 0.1 kg accuracy) were done while participants were wearing light clothes and no shoes. BMI was measured by
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squared body weight (kg) divided by height $^2$ (meter), fat percentage by Jackson/Pollock Caliper Method (28). Participants’ heart rate was measured and their aerobic capacity (VO$_{2}$max) by modified Bruce protocol (In the standard Bruce protocol, the starting point (ie, stage 1) is 1.7 mph at a 10% grade (5 METs). Stage 2 is 2.5 mph at a 12% grade (7 METs). Stage 3 is 3.4 mph at a 14% grade (9 METs). This protocol includes 3-minute periods to allow achievement of a steady state before workload is increased. The modified Bruce protocol has 2 warm up stages, each lasting 3 minutes. The first is at 1.7 mph and a 0% grade, and the second is at 1.7 mph and a 5% grade (29).

Exercise Program:

In the briefing sessions, participants were acquainted with research aims, the way exercises should be done and research time schedule.

**High Intensity Interval Training (HIIT):**

Participants exercised on treadmill for 8 weeks, 3 sessions in each weeks, with 90 percent of target heart rate (THR = Resting heart rate + Exercise intensity (Resting heart rate – Maximum heart rate)) (30).

The training program for each session included: 10 minutes for stretching and active warm-up, main exercise (repeating bouts for 4 minutes with 90 percent target heart rate and 2 minutes active recovery between bouts with 50-60 percent of target heart rate), and finally 5 minutes cool-down. These are shown in Table 1 (31). The heart rate of the participants were controlled. If increasing or decreasing exercise intensity was required, sufficient feedback was given to the participants.

<table>
<thead>
<tr>
<th>Table 1. HIIT program (3 sessions per week)</th>
</tr>
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<tbody>
<tr>
<td>weeks</td>
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<tr>
<td></td>
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<tr>
<td>1st week</td>
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<td>2nd week</td>
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<td>3rd week</td>
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<td>4th week</td>
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<td>5th week</td>
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<tr>
<td>6th week</td>
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<tr>
<td>7th week</td>
</tr>
<tr>
<td>8th week</td>
</tr>
</tbody>
</table>

Measuring Biochemical Variables:

Blood samples were taken after 12 hours of fasting and in 2 stages. In the first stage, according to the instructions, participants were asked to avoid doing any heavy physical activity, being in stressful conditions, and taking medicines 3 days prior to sampling. The resulting serums were frozen in -70℃ until the second experiment. The second sampling was done 48 hours after the last exercise session to remove the effects of this session from exercise and control groups. Serum amounts of FGF21 and Irisin were measured by ELISA Reader and Zellbio kit (made in Germany). Triglyceride, cholesterol, HDL-C and LDL-C were measured by photometric method using Pars Azmoon Co. kit.

Statistical Method:
The analysis of data was done using descriptive statistics for measuring central indexes and dispersion, K-S test for normal dispersion of data, leven test for determining the heterogeneity of variances, dependent t test for studying the pre-test and post-test changes made in dependent variables, and independent t test for studying differences between groups. Data were analyzed by SPPSS version 22 and significance level was p<0.05.

**Results**

Table 2 shows anthropometric and physiological characteristics of participants.

<table>
<thead>
<tr>
<th>Variable</th>
<th>group</th>
<th>Pre-test</th>
<th>Post-test</th>
<th>Inter-Group changes</th>
<th>Intra-group changes</th>
</tr>
</thead>
<tbody>
<tr>
<td></td>
<td></td>
<td></td>
<td></td>
<td>t</td>
<td>p</td>
</tr>
<tr>
<td>Age (years)</td>
<td>Control</td>
<td>30.5±2.27</td>
<td></td>
<td>0.531</td>
<td>0.608</td>
</tr>
<tr>
<td></td>
<td>Exercise</td>
<td>29.8±3.32</td>
<td></td>
<td></td>
<td></td>
</tr>
<tr>
<td>Height (cm)</td>
<td>Control</td>
<td>165.88±3.01</td>
<td></td>
<td>7.05</td>
<td># 0.001</td>
</tr>
<tr>
<td></td>
<td>Exercise</td>
<td>164.10±4.10</td>
<td></td>
<td></td>
<td></td>
</tr>
<tr>
<td>Body mass (kg)</td>
<td>control</td>
<td>83.05±5.99</td>
<td>82.88±5.41</td>
<td>0.608</td>
<td>1.140</td>
</tr>
<tr>
<td></td>
<td>Exercise</td>
<td>81.41±6.27</td>
<td>79.86±6.37</td>
<td></td>
<td></td>
</tr>
<tr>
<td>BMI (kg/m2)</td>
<td>control</td>
<td>30.24±1.19</td>
<td>30.02±1.36</td>
<td>0.604</td>
<td>0.143</td>
</tr>
<tr>
<td></td>
<td>Exercise</td>
<td>30.48±1.54</td>
<td>29.38±1.183</td>
<td></td>
<td></td>
</tr>
<tr>
<td>Fat (%)</td>
<td>control</td>
<td>41.26±3.35</td>
<td>40.86±3.25</td>
<td>0.840</td>
<td>0.432</td>
</tr>
<tr>
<td></td>
<td>Exercise</td>
<td>39.22±6.8</td>
<td>36.70±3.56</td>
<td></td>
<td></td>
</tr>
<tr>
<td>WHR (cm)</td>
<td>control</td>
<td>0.92±0.45</td>
<td>0.91±0.05</td>
<td>0.840</td>
<td>0.432</td>
</tr>
<tr>
<td></td>
<td>Exercise</td>
<td>0.91±0.33</td>
<td>0.87±0.20</td>
<td></td>
<td></td>
</tr>
<tr>
<td>VO2max (ml.kg.min-1)</td>
<td>Control</td>
<td>36.1±3.52</td>
<td>36.21±3.55</td>
<td>-1.129</td>
<td>0.288</td>
</tr>
<tr>
<td></td>
<td>Exercise</td>
<td>37.75±3.29</td>
<td>39.61±2.98</td>
<td></td>
<td></td>
</tr>
</tbody>
</table>

* Significance as compared to pre-test amounts
* Significance as compared to HIIT group. The amounts are indicated as standard deviation ± average.

Table 3 shows mean and standard deviation of dependent variables, before and after 8 weeks of exercise program. It also shows the results of dependent t test and independent t test.

The results of dependent t test showed that 8 weeks of HIIT increased serum amounts of Irisin (p=0.001) 7.87%, FGF21 (p=0.000) 6.11%, HDL-C (p=0.031) 7.97%, and VO2max (p=0.000) 4.92% in sedentary Obese women as compared with basic conditions. It also reduced weight (p=0.000) 1.90%, BMI (p=0.0007) 3.6%, body fat percentage (p=0.023) 6.42%, WHR (p=0.002) 4.93%, triglyceride (p=0.007) 4.31%, Total cholesterol (p=0.020) 3.03%, and CO/HDL (p=0.017) 10.22% as compared with basic conditions. No significant difference was observed in the control group (P>0.05). Also, the results of independent t test showed that there is a significant difference in Irisin (p=0.000), FGF21 (p=0.000), body fat percentage (p=0.014), WHR
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(p=0.014) and triglyceride (p=0.029) between HIIT and control groups. However, there was no significant difference between two groups in weight, BMI, VO₂max, HDL-C, LDL-C, cholesterol, LDL/HDL, and CO/HDL (p>0.05) (Table 3).

### Table 3. Results of Independent and Dependent t-statistic tests in determining the difference between research variables

<table>
<thead>
<tr>
<th>Variable</th>
<th>Group</th>
<th>Pre-test</th>
<th>Post-test</th>
<th>Inter-group changes</th>
<th>Intra-group changes</th>
</tr>
</thead>
<tbody>
<tr>
<td></td>
<td></td>
<td></td>
<td></td>
<td>t</td>
<td>p</td>
</tr>
<tr>
<td>Irisin (ng/ml)</td>
<td>Control</td>
<td>162.03±2.45</td>
<td>162.29±4.36</td>
<td>-0.147</td>
<td>0.865</td>
</tr>
<tr>
<td></td>
<td>Exercise</td>
<td>162.73±4.45</td>
<td>175.55±7.22</td>
<td>-4.692</td>
<td># 0.001</td>
</tr>
<tr>
<td>FGF21 (pg/ml)</td>
<td>Control</td>
<td>249.73±2.67</td>
<td>249.90±3.35</td>
<td>-0.141</td>
<td>0.891</td>
</tr>
<tr>
<td></td>
<td>exercise</td>
<td>250.85±4.44</td>
<td>265.93±6.62</td>
<td>-6.232</td>
<td># 0.000</td>
</tr>
<tr>
<td>WHR (cm)</td>
<td>Control</td>
<td>0.92±0.45</td>
<td>0.91±0.05</td>
<td>0.840</td>
<td>0.432</td>
</tr>
<tr>
<td></td>
<td>exercise</td>
<td>0.91±0.33</td>
<td>0.87±0.20</td>
<td>4.260</td>
<td># 0.0002</td>
</tr>
<tr>
<td>VO₂max (ml.kg.min⁻¹)</td>
<td>Control</td>
<td>36.1±3.52</td>
<td>36.21±3.55</td>
<td>-1.129</td>
<td>0.288</td>
</tr>
<tr>
<td></td>
<td>Exercise</td>
<td>37.75±3.29</td>
<td>39.61±2.98</td>
<td>-6.811</td>
<td># 0.000</td>
</tr>
<tr>
<td>Triglyceride (mg/dl)</td>
<td>Control</td>
<td>128.92±4.73</td>
<td>127.93±3.87</td>
<td>1.371</td>
<td>0.204</td>
</tr>
<tr>
<td></td>
<td>exercise</td>
<td>129.56±3.63</td>
<td>129.56±3.63</td>
<td>3.475</td>
<td># 0.007</td>
</tr>
<tr>
<td>Total cholesterol (mg/dl)</td>
<td>Control</td>
<td>169.46±5.31</td>
<td>168.64±4.31</td>
<td>1.155</td>
<td>0.287</td>
</tr>
<tr>
<td></td>
<td>exercise</td>
<td>170.94±3.28</td>
<td>165.76±4.96</td>
<td>2.827</td>
<td># 0.020</td>
</tr>
<tr>
<td>LDL-C (mg/dl)</td>
<td>Control</td>
<td>103.56±12.04</td>
<td>103.21±13.89</td>
<td>0.210</td>
<td>0.839</td>
</tr>
<tr>
<td></td>
<td>exercise</td>
<td>102.28±7.149</td>
<td>95.40±12.15</td>
<td>1.594</td>
<td>0.146</td>
</tr>
<tr>
<td>HDL-C (mg/dl)</td>
<td>Control</td>
<td>43.91±4.03</td>
<td>44.64±4.72</td>
<td>-1.017</td>
<td>0.335</td>
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<tr>
<td></td>
<td>exercise</td>
<td>42.99±4.82</td>
<td>46.42±4.80</td>
<td>-2.55</td>
<td># 0.031</td>
</tr>
<tr>
<td>CO/HDL (mg/dl)</td>
<td>Control</td>
<td>3.87±0.368</td>
<td>3.81±0.393</td>
<td>0.958</td>
<td>0.363</td>
</tr>
<tr>
<td></td>
<td>Exercise</td>
<td>4.01±0.44</td>
<td>3.60±0.452</td>
<td>2.913</td>
<td># 0.017</td>
</tr>
<tr>
<td>LDL-C/HDL-C (mg/dl)</td>
<td>Control</td>
<td>2.32±0.45</td>
<td>20.25±0.411</td>
<td>0.510</td>
<td>0.622</td>
</tr>
<tr>
<td></td>
<td>exercise</td>
<td>2.48±0.43</td>
<td>2.12±0.54</td>
<td>1.44</td>
<td>0.182</td>
</tr>
</tbody>
</table>

* Significance as compared to pre-test amounts

* Significance as compared to HIIT group. The amounts are indicated as standard deviation ± average.

### Discussion

The importance of exercise in preventing and curing metabolic and cardiovascular diseases is clear today. However, cellular-molecular and other mechanisms through which these activities exert their positive influence are not fully comprehended yet. Different studies have been done on brown adipose tissue as the effective tissue in energy hemostasis in recent years. These studies show that FGF21 and Irisin create heat, energy expenditure, and prevent obesity by transforming WAT to BAT and activating UCP1 (27). Thus, the aim of this research is to investigate the effect of eight weeks of high intensity interval training (HIIT) on serum amounts of FGF21 and Irisin in obese, sedentary women.
The results showed that 8 weeks of HIIT exercise increased serum amounts of FGF21 in obese, sedentary women significantly and there was a significant difference between two groups. There are limited and contradictory studies about the effect of HIIT on FGF21. Dlouhy et al. (2016) showed that 8 weeks of aerobic exercises with 60 to 75% maximum heart rate for 30 to 45 minutes increase FGF21 serum amounts significantly but there was no significant change in the control group (32). Cuevas et al. (2012) showed that 2 weeks of aerobic training for 5 days a week can increase FGF21 serum amounts in healthy women (33). Kong et al. (2016) showed that 5 weeks of HIIT training including sixty 8-second repetitions with 12-second intervals, didn’t make a significant difference in serum amounts of FGF21 in obese women (16). Also, Birjandi et al. (2016) showed that 6 weeks of HIIT (3 sessions per week) didn’t make a significant difference in serum amounts of FGF21 in obese women (17). The existing differences can be due to the difference in groups, number of participants, intensity and duration of the exercise, gender, age, and the time between the last exercise session and blood sampling. For instance, in Slusher et al. (2015) study, blood sampling was done right after the last exercise session while in this research it was done 48 hours after the last session (14). One can compare the study by Kong (5 weeks of training) with that of Birjandi (6 weeks of training) about the effect of exercise duration on the results (16,17).

Mechanisms through which HIIT can increase FGF21 are unknown but the following mechanisms are suggested for justifying increase in FGF21 and its physiologic effects:

AMPK’s activation while doing intensive exercise can be one of the factors. AMPK is the key modifier for fat metabolism and energy balance. AMPK gets activated with the increase in inter-cellular AMP/ATP and its activation leads to the activation of metabolic pathway producing ATP (33). Thus, AMPK’s activation activates peroxisome proliferator-activated receptor alpha (PPARα) and accordingly increases FGF21 (33).

Glucagon ration to insulin is considered FGF21 modifier and glucagon’s content increases by sport activity (34). Insulin motivates FGF21 in skeleton muscles. Muscular FGF21 increases significantly a few hours after insulin injection. Insulin increases FGF21 by activating PI3/AKT signaling pathway. In addition to insulin, muscular contraction activates AKT signaling pathway and increases muscular FGF21. Therefore, insulin and skeletal muscle contraction are two main activators of AKT in skeletal muscle. During the training, insulin decreases but skeletal muscle contraction continues which can be a mechanism for activating AKT pathway independent from insulin (15).

It is shown that FGF21 amounts have a direct link with BMI, visceral adiposity, and pericardial adiposity (35). Also, there is a direct link between FGF21 and physical fitness. So, those with low physical fitness have low amounts of FGF21 (36). As a result, HIIT training in this research increases FGF21 by activating AMPK, PI3K/AKT, and increasing VO_{2}max.

FGF21 shows its anti-obesity effects in the following mechanisms:

FGF21 control the body weight by activating mTORC1 and increasing the absorption of glucose, oxidation of free fatty acids and glycolysis (15).

FGF21 increases insulin signaling significantly. It also increases glucose absorption by releasing adipokines sensitive to insulin like adiponectin. In fact, FGF21 increases adiponectin and adiponectin increases β-oxidation though ceramide-adiponectin-FGF21. It also increases energy consumption, decreases glucose, decreases ceramide acid in liver and adipose tissue, improves insulin sensitivity and reduces weight (37).

FGF21 controls liver lipogenesis and increases FFA lipolysis when the person is hungry (38). In response to hunger, FFA is used by activating free fatty acids oxidation and lipolysis in liver tissue (33). Similar to
hunger, in exercise training lipolysis and FFA oxidation are increased as main fuels for ATP production in liver and skeletal muscles tissues and FFA increases the activity of PPARα in the liver which leads to FGF21 increasing (33).

FGF21 increases energy consumption by modifying AMPK activity and SIRT1 through LKB1 in adipose tissue. Change in NAD⁺ amounts in a cell activates SIRT1, PGC-1α, and histone-3. Their activation increases the function of mitochondrial oxidative and reduces weight (11).

One of the other effective FGF21 mechanisms is the conversion of white adipose tissue to brown one (33). FGF21 increases UCP1 and other heat producing genes in adipose tissue. An increase in PGC-1α protein modifies this process. PGC-1α produces heat by activating UCP1 in the brown adipose tissue (39). In this research, HIIT reduced fat mass, BMI and WHR. Although FFA and UCP1 were not measured in this research, it seems fat mass reduction is an indirect index for FFA increase which shows lipolysis of fat tissue.

Also, the results of this research showed that 8 weeks of HIIT can increase serum amounts of Irisin in obese, sedentary women significantly. There was a significant difference between HIIT and control groups. These results support those of the study done by Molaei et al. (2015) and Khalafi et al. (2016) which showed the increase in irisin in rats after 10 weeks of HIIT and its increase after 8 weeks of HIIT and aerobic training with low intensity respectively (40,41). Miyamoto et al. (2015) showed that there is no significant difference in serum amounts of irisin in professional athletes, occasional athletes, and sedentary people (44). Hakimi et al. (2015) showed that 8 weeks of aerobic training with 60 to 80 percent of heart rate in male overweight university students didn’t make any changes in their Irisin serum levels (42).

Irisin is a hormone motivated by exercise and secreted in mice and human beings by skeletal muscle. It stimulates white adipose tissue conversion to brown adipose tissue which improves energy consumption rate (18). In this research, it seems AMPK’s activation during HIIT training is one of the factors for increasing PGC-1α and irisin. AMPK’s activation leads to the phosphorylation of PGC-1α as FNDC5’s modifier and irisin secretion (43). Also, PGC-1α activates PPARγ. PPARγ is involved in energy metabolism and stimulates FNDC5 and irisin increase (44). It is shown that there is a relation between irisin amounts and precursor of FNDC5 and PGC-1α (43). Gene expression of FNDC5 in skeletal muscle is related to obesity in human. The amount of expression increases with obesity. This relation can be a compensatory mechanism (18,45). In line with this theory, another research showed gene expression of FNDC5 in skeletal muscle has a positive relation with BMI (18,45). Some studies reveal that irisin has a reverse relation with BMI, glucose and triglyceride (46). Also, some studies show that irisin amounts have a positive correlation with muscular mass and a negative correlation with fat mass (47).

Obesity comes with metabolic disturbances such as increase in cholesterol serum amount, LDL-C, VLDL-C, TG, and decreasing HDL (47). The results of this research didn’t showed a significant reduction in LDL-
C and HDL/LDL but showed significant reduction in TG, total cholesterol, cholesterol on HDL, BMI, WHR, and fat percentage. It also showed significant increase in HDL-C amounts. In this regard, Elmer et al. (2016) showed 8 weeks of HIIT compared with moderate intensity aerobic training increased TG but didn’t change HDL-C and cholesterol significantly (44). Peterson et al. (2015) showed that 6 weeks of HIIT (each session including 2 thirty-minute intervals of indoor cycling) decreased LDL, cholesterol, and TG while change in HDL was not significant (*). Hassani et al. (2016) showed that 8 weeks of HIIT deceased TG significantly but made no significant change in cholesterol, LDL, and HDL (55). Paahoo. (2015) showed that 12 weeks of HIIT decreased GT, LDL-C and increased HDL-C in sedentary obese boys significantly (56). The differences can be due to the difference in the used HIIT protocols. Using different durations, intensities, and types of HIIT make it possible to create unlimited number of protocols.

An intensity that optimizes fat oxidation as the main energy source during exercise is important. Research has shown that obese individuals have an impaired utilization of free fatty acid in the skeletal muscle (57). Endurance training is an effective strategy for obesity prevention and weight loss because it enhances lipolysis and fatty acid oxidation in the skeletal muscle. This enhancement is known to be intensity-dependent, as the absolute rate of fat oxidation (g.min-1) increases from low to moderate intensity and then decreases as exercise becomes more intense (58). Nevertheless, studies show six weeks of HIIT for 2 weeks can increase skeletal muscle oxidative capacity, endurance performance and change metabolic amount (54). Storina et al. (2013), showed that 2 weeks of HIIT (6 sessions of 4 to 6 repeats of 30s Wingate with 4-5 min recovery) increased resting fat oxidation in overweight/obese sedentary men (59).

Mechanisms through which HIIT leads to increase in fat oxidation and losing weight are unknown. However, it seems HIIT training in this research leads to fat and weight loss and fat oxidation through the following mechanisms:

It seems that fatty acid transport proteins have been linked to enhanced fat oxidation. The increases in fatty acid translocase (FAT/CD36) and plasma membrane fatty acid-binding protein (FABPpm) both found in the sarcolemma, the mitochondrial membrane, and in a cytoplasmic pool in skeletal muscle, could contribute to the enhanced fat oxidation by increasing the rate of free fatty acid transfer across the muscle and mitochondrial membrane (54). It has been shown that Six weeks of HIIT (ten 4-min cycling bouts at 90% VO2peak separated by 2-min of rest) increased fatty acid transport protein content in whole muscle (FAT/CD36 and FABPpm), sarcolemmal (FABPpm) and mitochondrial (FAT/CD36) membranes in the skeletal muscle of 10 untrained females, suggesting that increases in skeletal muscle fatty acid oxidation following exercise training at high intensity are related in part to changes in fatty acid transport protein content (56).

Exercises training increases lipoprotein type A and LPL, a key enzyme in fat metabolism. The increase in LPL activity can be due to increase in catecholamine, discharge of energy loads, increase in free radicals, and decrease in cellular PH (57). Although LPL activity is not measured in this study, it is probable that its increase has a reverse relation with TG which leads to TG breaking (57). Also, LPL has a direct relation with HDL and leads to the evolution of HDL molecules (58). Moreover, HDL increase can be attributed to factors such as LCAT increase (59). It seems that HIIT increases LPL enzyme, LCAT and HDL activity but decreases LDL, TG, and cholesterol (59).

Exercise training can be an important intervention for losing weight. Exercise represents an important intervention for weight loss as it has the potential to
reduce body mass, increase fat-free mass, and maintain or elevate resting metabolic rate (55). A number of studies have demonstrated that HIIT may induce weight loss in sedentary overweight/obese individuals. For example, a significant reduction in waist circumference and subcutaneous adipose tissue was found after 2 weeks of HIIT in overweight/obese sedentary men (55).

Although the mechanism responsible for fat and weight loss after HIIT is unclear, one possible reason is an increase in post-exercise metabolism (60). Excess post-exercise oxygen consumption (EPOC) response to HIIT may have a role in elevating post-exercise fat oxidation through the increased levels of catecholamine generated during acute HIIT (60). Increase in plasma epinephrine and norepinephrine at the end of HIIT could increase lipolysis and the availability of free fatty acids, resulting in increased overall fat oxidation during and after HIIT (61). Moreover, HIIT significantly increases muscle mitochondrial beta-hydroxacyl-CoA dehydrogenase, which may enhance fat loss (61). Also, the need to remove lactate and H+ and to resynthesize glycogen during and after HIIT also increases fat oxidation (53).

Decreased post-exercise appetite is another possible mechanism underlying HIIT-induced fat loss. Although the effect of HIIT on appetite suppression has not been investigated in overweight/obese individuals, a single bout of intense exercise has been found to suppress hunger immediately following cessation of the exercise (53). In investigating the effects of HIIT (6×30s Wingate tests) and endurance exercise (60 min exercise at 68.1% of VO₂max) on appetite. The men reported higher appetite perceptions in the hours after an acute bout of HIIT than after the endurance exercise (62). There is no clear mechanism that explains why hunger level is suppressed after high-intensity exercise. However, there is evidence to suggest a marked effect of intense exercise on subjective hunger based on the reports of exercise-induced anorexia (53). Also, decreasing appetite after HIIT can be explained through by the considerable redistribution of blood flow away from the splanchnic circulation into the working muscles (53). While acute exercise reduces liver and muscle glycogen stores, which may result in an immediate increase in hunger, chronic exercise training may induce more adaptations that may lead to more stable levels of metabolic fuels, resulting in a suppression of hunger (63). In summary, weight loss and fat oxidation resulting from HIIT can be explained by lipolytic enzymes, fatty acid transport proteins (FAT/CD36, FABPpm, appetite decrease and negative energy balance through EPOC.

Different compatibility responses of individuals to the exercise, lack of some participants’ control over their emotions, sleep rate, and lack of control on participants’ foods are among the limitations of this study.

To sum up, this study shows that HIIT can cause an increase in FGF21 and Irisin and thus transform white adipose tissue to brown adipose tissue and preventing obesity and its negative consequences. Therefore, it is suggested that obese individuals take part in HIIT - As a time-efficient training method - and overcome their obesity-related problems.

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